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## INTERNATIONAL PROTEOLYSIS SOCIETY OUCCKCUTS Editors: Sheena McGowan (Monash University)



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#### THE PREMIER RESOURCE FOR ALL YOUR IMPORTANT PROTEASE QUESTIONS

ANUARY 2017

Margarete Heck (University of Edinburgh)

## A Message From the President:

Protease research and other subjects of biochemistry are increasingly seeing competition by areas of the life sciences that are just growing bigger and hotter (some people would say more "sexy") every year. But still – regulated proteolysis is a fundamental principle of normal homeostasis ad deregulated proteolysis is a hallmark of many disease conditions.

Certainly our readers would support this statement – but how to convince junior scientists to stay in the field after completing their Ph.D. or postdoc? IPS has a long-standing commitment on encouragement, training, and support of early carrier scientists at the biannual general meetings of the society. Recently we extended our outreach by the IPS council decision to support also other protease-related meetings by rather small amounts of money. In this regard, the first action was to sponsor the meeting "Metalloproteinases and their inhibitors: beginning, past & future" held at Keble College Oxford in August 2016. At this meeting, organized by Linda Troeberg (Oxford), Jelena Gavrilovic (UEA), and Yoshifumi Itoh (Oxford), IPS sponsored an oral presentation award to Simone D. Scilabra (TU Munich) and two poster prizes to Pernille Søgaard (University of Oxford) and Kim Lemmens (KU Leuven) Congratulations! If you plan to organize a protease meeting and still need some support – just contact us at ipssecretary@gmail.com.

In this issue of Quick Cuts we advertise our 10th General Meeting of the Society held in beautiful Banff, Canada, from Oct. 28th- Nov. 2nd, 2017. I am sure - Joanne Lemieux, Jean-Bernard Denault and Christopher Overall are preparing an outstanding meeting at the frontiers of proteolysis. IPS will help the meeting by providing support for travel awards and training workshops. Our support of 'members in training' relies on your support of the society. Please renew your IPS membership!! For the majority of IPS members the renewal will be due in January 2017.

We are also inviting nominations for honorary Lifetime Members of the IPS to be presented at the Banff meeting. Candidates should have a track record of accomplishments in the protease world and be friends of IPS. Please send names and a short case for support to Ulrich auf dem Keller (ipssecretary@gmail.com). A list of our Lifetime Members can be found at http://prote-ase.org/LifetimeMembers.html.

Finally thanks to Sheena McGowan and Margarete Heck for preparing this issue of Quick Cuts. Enjoy reading! Thomas Reinheckel, IPS President

## MEETING REPORT Metalloproteinases and their inhibitors: *beginning, past and future.* 4-5 August 2016, Keble College Oxford UK

This conference was held at Keble College, Oxford to the contribution of two leading figures in field, Professors Hideaki Nagase (University of Oxford) and Professor Gillian Murphy (University of Cambridge) upon their retirements and 70<sup>th</sup> birthdays. The meeting was co-organised by Yoshifumi Itoh (University of Oxford), Linda Troeberg (University of Oxford) and Jelena Gavrilovic (University of East Anglia), who have worked closely with Hideaki and Gill for many years. 100 delegates attended the meeting, including 14 invited speakers, 33 principal investigators and more than 40 graduate students and post-doctoral researchers. The meeting was international as invited speakers were from Canada, USA, Japan, Germany, Israel, Italy, and UK, and delegates were from USA, Brazil, Japan, Israel, Denmark, Finland, Belgium, Germany, Italy and UK.

Metalloproteinases are a large group of proteolytic enzymes that modify the microenvironment of cells and play crucial roles in tissue remodeling. Talks and posters covered a broad range of topics, reflecting the important role metalloproteinases playing in physiological processes such as development and immunity and also in pathophysiological settings such as cancer and arthritis. Evaluation of the enzymes as potential therapeutic targets or tools for diagnosis were highlighted by several speakers. Keynote presentations by Professors Nagase and Murphy stressed that understanding the fundamental biochemistry of metalloproteinases is a prerequisite for reaching their translational potential. An emerging theme was the role of metalloproteinases in subtle modulation of protein function. For example, Christopher Overall (University of British Columbia) and William C. Parks (Cedars-Sinai Medical Center) discussed how metalloproteinases regulate immune responses by processing cytokines, chemokines and matrix proteins.

Young scientists were well represented at the meeting, with 8 speakers selected from submitted abstracts. Simone D. Scilabra (Technische Universität München, Germany) won the oral presentation prize for his talk on development of a 'trap' to increase levels of the protective metalloproteinase inhibitor TIMP-3 in tissue. Pernille Søgaard (University of Oxford) and Kim Lemmens (KU Leuven) won the poster presentation prizes, for their work on the collagen receptor DDR1 and on axonal regeneration in zebrafish, respectively.



Feedbacks from delegates have been overwhelmingly positive, with attendees enjoying the opportunity to come together and celebrate Professors Nagase and Murphy's contribution to our field. Students were particularly benefited from the strong line-up of worldleading international speakers. The meeting provided the community with a valuable opportunity to reflect on the history of this field and to identify future research priorities.

Linda Troeberg Yoshifumi Itoh University of Oxford



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ociety

## **IMPORTANT PROTEASE PAPERS I**

#### **Research Publications**

#### **AGONIST & INHIBITORS**

Drinkwater, N., Vinh, VB., Mistry, SN., Bamert, RS., Ruggeri, C., Holleran, JP., Loganathan, S., Paiardini, A. Charman, SA., Powell, AK., Avery, VM., McGowan, S. & Scammells, PJ.

Potent dual inhibitors of *Plasmodium falciparum* M1 and M17 aminopeptidases through optimization of S1 pocket interactions.

Eur. J. Med. Chem. 2016. 110:43-64.

Ruggeri, C., Drinkwater, N., Kannan Sivaraman, K., Bamert, RS., McGowan, S. & Paiardini, A.

Identification and validation of a potent dual inhibitor of the *Plasmodium falciparum* M1 and M17 aminopeptidases using virtual screening.

PLOS ONE. 2015. 10(9):e0138957,

Lieu T, Savage E, Zhao P, Edgington-Mitchell L, Barlow N, Bron R, Poole DP, McLean P, Lohman R-J, Fairlie DP, and Bunnett NW.

Antagonism of the proinflammatory and pronociceptive actions of canonical and biased agonists of protease-activated receptor-2.

British J of Pharmacol. 2016. 173: 2752-2765.

Weider E, Susan-Resiga D, Essalmani R, Asselin M.-C, Hamelin J, Marcinkiewicz J, Nimesh N, Wycoff KL, Zhang J, Prat A, and Seidah NG.

Single domain antibodies as potent inhibitors of PCSK9 activity on LDLR degradation.

J Biol Chem. 2016. 291:16659-16671.

Niemiec E, Chevrier F, Roy V, Garenne T, Lecaille F, Warszycki D, Bojarski AJ, Lalmanach G, and Agrofoglio LA

Straightforward synthesis of 2,4,6-trisubstituted 1,3,5-triazine compounds targeting cysteine cathepsins K and S

Eur. J. Med. Chem. 2016. 12: 12-20.

Cleary, JA.& Malthouse, JPG.

A new Lysine derived glyoxal inhibitor of trypsin, its properties and utilization for studying the Stabilization of Tetrahedral adducts by Trypsin.

Biochem Biophys Rep 2016. 5: 272-284.

Cleary JA, Doherty W, Evans P, Malthouse JP.

Quantifying tetrahedral adduct formation and stabilization in the cysteine and the serine proteases.

Biochim Biophys Acta (BBA) - Proteins & Proteomics 2015. 1854: 1382-1391.

#### **PROTEASE PROBES**

Tamhane T, Wolters BK, Illukkumbura R, Maelandsmo GM, Haugen MH, and Brix K.

Construction of a plasmid coding for green fluorescent protein tagged cathepsin L and data on expression in colorectal carcinoma cells.

Data in Brief. 2015. 5: 468-475.

Poreba M, Solberg R, Rut W, Lunde NN, Kasperkiewicz P, Snipas SJ, Mihelic M, Turk D, Turk B, Salvesen GS, & Drag M.

Counter Selection Substrate Library Strategy for Developing Specific Protease Substrates and Probes.

Cell Chem Biol. 2016. 23: 1- 13.

#### **MICROBIAL PROTEASES**

Hwang J, Ribbens D, Raychaudhuri S, Cairns L, Gu H, Frost A, Urban S & Espenshade PJ.

A Golgi rhomboid protease Rbd2 recruits Cdc48 to cleave yeast SREBP.

EMBO Journal. 2016. In Press

Liu P, Robbins AH, Marzahn MR, McClung SH, Yowell CA, Stevens Jr. SM, Dame JB, and Dunn BM.

Enzymatic characterization of recombinant food vacuole plasmepsin 4 from the rodent malaria parasite *Plasmodium berghei*.

PLoS ONE. 2015. 10: e0141758.

Hammoudi PM, Jacot D, Mueller C, Di Cristina M, Dogga SK, Marq JB, Romano J, Tosetti N, Dubrot J, Emre Y, Lunghi M, Coppens I, Yamamoto M, Sojka D, Pino P, Soldati-Favre D.

Fundamental Roles of the Golgi-Associated Toxoplasma Aspartyl Protease, ASP5, at the Host-Parasite Interface.

PLoS Pathog. 2015. 11.

Dvořák J, Fajtová P, Ulrychová L, Leontovyč A, Rojo-Arreola L, Suzuki BM, Horn M, Mareš M, Craik CS, Caffrey CR, O'Donoghue AJ.

Excretion/secretion products from *Schistosoma mansoni* adults, eggs and schistosomula have unique peptidase specificity profiles.

Biochimie. 2016. 122:99-109

Sojka D, Pytelková J, Perner J, Horn M, Konvičková J, Schrenková J, Mareš M, Kopáček P.



## **IMPORTANT PROTEASE PAPERS II**

Multienzyme degradation of host serum albumin in ticks.

Ticks Tick Borne Dis. 2016, 7, 604-613.

LaRock CN, Todd J, LaRock DL, Olson J, O'Donoghue AJ, Robertson AAB, Cooper MA, Hoffman HM, Nizet V.

IL-1 $\beta$  is an innate immune sensor of microbial proteolysis

Science Immun. 2016. 1(2):pp. eaah3539

Goupil LS, Ivry SL, Hsieh I, Suzuki BM, Craik CS, O'Donoghue AJ, McKerrow JH.

Cysteine and Aspartyl Proteases Contribute to Protein Digestion in the Gut of Freshwater Planaria.

PLoS Negl Trop Dis. 2016. 10(8)e0004893

Naumann TA, Naldrett MJ, Ward TJ, and Price NPJ.

Polyglycine hydrolases: Fungal beta-lactamase-like endoproteases that cleave polyglycine regions within plant class IV chitinases.

Protein Science. 2015. 24: 1147-1157.

Bailleul A, Kravtzoff A, Joulin-Giet A, Lecaille F, Labas V, Loth K, Meudal H, Texeira-Gomes AP, Gilbert FB, Coquet L, Jouenne T, Brömme D, Schouler C, Landon C, Lalmanach G, and Lalmanach AC.

The unusual resistance of avian defensin AvBD7 to proteolytic enzymes preserves its antibacterial activity

PLoS One. 2016. 11(8): e0161573

Almeida CM, Gomes D, Faro C, Simões I.

Engineering a cardosin B-derived rennet for sheep and goat cheese manufacture.

Appl Microbiol Biotechnol. 2015 Jan;99(1):269-81.

Curto P, Simões I, Riley SP, Martinez JJ

Differences in Intracellular Fate of Two Spotted Fever Group Rickettsia in Macrophage-Like Cells

Front. Cell. Infect. Microbiol. 2016 6:80

Leal AR, Cruz R, Bur D, Huesgen PF, Faro R, Manadas B, Wlodawer A, Faro C, Simões I

Enzymatic properties, evidence for in vivo expression, and intracellular localization of shewasin D, the pepsin homolog from Shewanella denitrificans.

Sci Rep. 2016 Mar 31;6:23869.

#### **PROTEASES & PATHOLOGIES**

O'Donoghue AJ, Ivry SL, Chaudhury C, Hostetter DR, Hanahan D, Craik CS.

Procathepsin E is highly abundant but minimally active in pancreatic ductal adenocarcinoma tumors.

#### Biol Chem. 2016. 397(9):871-81

Haugen MH, Boye K, Nesland JM, Pettersen SJ, Egeland EV, Tamhane T, Brix K, Maelandsmo GM, and Flatmark K.

High expression of the cysteine proteinase legumain in colorectal cancer - Implications for therapeutic targeting.

Eur J Cancer. 2015. 51: 9-17.

Guillemot J, Asselin M-C, Susa-Resiga D, and Seidah NG.

Deferoxamine stimulates LDLR expression and LDL uptake in HepG2 cells.

Mol. Nutr. Food. Res. 2016. 60:600-608.

Tamhane T, Illukkumbura R, Lu S, Maelandsmo GM, Haugen MH, and Brix K.

Nuclear cathepsin L activity is required for cell cycle progression of colorectal carcinoma cells.

Biochimie. 2016. 122: 208-218.

Fuhrman-Luck RA, Stansfield SH, Stephens CR, Loessner D, Clements JA.

Prostate Cancer-Associated Kallikrein-Related Peptidase 4 Activates Matrix Metalloproteinase-1 and Thrombospondin-1.

J Proteome Res. 2016. 15:2466-2478.

Dorn J, Yassouridis A, Walch A, Diamandis EP, Schmitt M, Kiechle M, Wang P, Drecoll E, Schmalfeldt B, Loessner D, Kotzsch M, Magdolen V.

Assessment of kallikrein-related peptidase 5 (KLK5) protein expression in tumor tissue of advanced ovarian cancer patients by immunohistochemistry and ELISA: correlation with clinical outcome.

Am J Cancer Res. 2015. 6:61-70.

Lisle JE, Mertens-Walker I, Stephens CR, Stansfield SH, Clements JA, Herington AC, Stephenson SA.

Murine, but not human, ephrin-B2 can be efficiently cleaved by the serine protease kallikrein-4: Implications for xenograft models of human prostate cancer.

Exp Cell Res. 2015. 333:136-46.

Paschkowsky S, Hamzé M, Oestereich, F, Munter LM.

Alternative Processing of the Amyloid Precursor Protein Family by Rhomboid Protease RHBDL4.

J Biol Chem 2016. In Press

Wong PF, Gall MG, Bachovchin WW, McCaughan GW, Keane FM, and Gorrell MD.

## **IMPORTANT PROTEASE PAPERS III**

Neuropeptide Y is a physiological substrate of fibroblast activation protein: Enzyme kinetics in blood plasma and expression of Y2R and Y5R in human liver cirrhosis and hepatocellular carcinoma.

Peptides 2016. 75: 80-95.

Hillebrand LE, Bengsch F, Hochrein J, Hülsdünker J, Bender J, Follo M, Busch H, Boerries M, and Reinheckel T.

Proteolysis - a characteristic of tumor-initiating cells in murine metastatic breast cancer.

Oncotarget. 2016. In Press

Öhrvik H, Logeman B, Turk B, Reinheckel T, and Thiele DJ.

Cathepsin Protease Controls Copper and Cisplatin Accumulation via Cleavage of the Ctr1 Metal-binding Ectodomain.

J Biol Chem. 2016. 291:13905-13916.

#### **PROTEOMICS & SYSTEMS BIOLOGY**

Huesgen PF, Lange PF, Rogers LD, Solis N, Eckhard U, Kleifeld O, Goulas T, Gomis-Rüth FX and Overall CM.

LysargiNase mirrors trypsin for identification of protein Ctermini and methylation sites.

Nat. Meth. 2015. 12: 55-58.

Sabino F, Hermes O, Egli FE, Kockmann T, Schlage P, Croizat, P, Kizhakkedathu JN, Smola H, and auf dem Keller U.

*In vivo* assessment of protease dynamics in cutaneous wound healing by degradomics analysis of porcine wound exudates.

Mol Cell Proteomics. 2015. 14: 354-370.

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Monitoring matrix metalloproteinase activity at the epidermaldermal interface by SILAC-iTRAQ-TAILS.

Proteomics. 2015. 15: 2491-2502.

Prudova A, Gocheva V, auf dem Keller U, Eckhard U, Olson OC, Akkari L, Butler GS, Fortelny N, Lange PF, Mark J, Joyce J, and Overall CM.

TAILS N-terminomics and Proteomics Show Protein Degradation Dominates Over Proteolytic Processing by Cathepsins in Pancreatic Tumors.

Cell Rep. 2016. 16: 1762-1773.

#### STRUCTURE

Porebski, B., Keleher, S., Hollins, JJ., Nickson, AA., Marijanovic, EM., Borg, NA., Costa, MGS., Pearce, MA., Dai, W., Zhu, L., Irving, J., Hoke, DE., Kass, I., Whisstock, JC., Bottomley, SP., Webb, GI., McGowan, S. & Buckle, AM.

Smoothing a rugged protein folding landscape by sequencebased redesign.

Nat Sci Rep. 2015. 6: 33958.

Drinkwater, N., Kannan Sivaraman K., Bamert, RS., Rut, W., Mohamed, Vinh, N., Scammells, PJ., Drag M & McGowan, S.

Structure and Substrate Fingerprint of Aminopeptidase P from *Plasmodium falciparum*.

Biochem J. 2016. 473:3189-3204

Garcia-Ferrer I, Arêde P, Gómez-Blanco J, Luque D, Duquerroy S, Castón JR, Goulas T and Gomis-Rüth FX.

Structural and functional insights into Escherichia coli  $\alpha_2$ -macroglobulin endopeptidase snap-trap inhibition.

Proc. Natl. Acad. Sci. USA. 2015. 112: 8290-8295.

Riley, BT, Ilyichova O, Costa MG, Porebski BT, de Veer SJ, Swedberg JE, Kass I, Harris JM, Hoke DE, Buckle AM.

Direct and indirect mechanisms of KLK4 inhibition revealed by structure and dynamics

Sci Rep. 2016. In Press.

Arolas JL, Goulas T, Pomerantsev AP, Leppla SH and Gomis-Rüth FX.

Structural basis for latency and function of immune inhibitor A metallopeptidase, a modulator of the *Bacillus anthracis* secretome.

Structure. 2016. 24: 25-36.

Masuyer G, Douglas RG, Sturrock ED, Acharya KR.

Structural basis of Ac-SDKP hydrolysis by angiotensin-1converting enzyme

Sci Rep. 2015. 5:13742-13752.

Cho S, Dickey SW, and Urban S.

Crystal structures and inhibition kinetics reveal a two-stage catalytic mechanism with drug design implications for rhomboid proteolysis.

Molecular Cell. 2016. 61: 329-340.

Larmuth KM, Masuyer G, Douglas RG, Schwager SL, Acharya KR, and Sturrock ED.

Kinetic and structural characterization of amyloid-β peptide hydrolysis by human angiotensin-1-converting enzyme.

## **IMPORTANT PROTEASE PAPERS IV**

#### FEBS J. 2016. 283(6):1060-76.

Kristensen LH, Olsen OH, Blouse GE, Brandstetter H.

Releasing the brakes in coagulation Factor IXa by cooperative maturation of the substrate-binding site.

Biochem J. 2016. 473(15):2395-411

Li M, Gustchina A, Cruz R, Simões M, Curto P, Martinez J, Faro C, Simões I, Wlodawer A.

Structure of RC1339/APRc from *Rickettsia conorii,* a retropepsin-like aspartic protease.

Acta Crystallogr D Biol Crystallogr. 2015 (Pt 10):2109-18.

#### ADAMS

Madoux, Pettiloud, Santos, Becker-Pauly, Fields, Bannister, Spicer, Cudic, Scampavia, and Minond.

Discovery of a selective, time-dependent inhibitor of ADAM10 using an exosite-binding glycosylated substrate.

#### Sci Rep 2016. In Press

Knapinska, Dreymuller, Smith, Golubkov, Fridman, Giulianotti, Houghten, Fields and Minond.

SAR Studies and Characterization of Exosite Inhibitors of A Disintegrin And Metalloprotease 17 (ADAM17) as In Vitro Biological Probes.

J Med Chem. 2015. 58: 5808-24.

Moss ML, Minond D, Yoneyama T, Hansen HP, Vujanovic N, Rasmussen FH.

An improved fluorescent substrate for assaying soluble and membrane-associated ADAM family member activities.

Anal Biochem 2016 507: 13-7.

#### CATHEPSINS

Oresic Bender K, Ofori L, van der Linden WA, Mock ED, Datta GK, Chowdhury S, Li H, Segal E, Sanchez Lopez M, Ellman JA, Figdor CG, Bogyo M, and Verdoes M.

Design of a highly selective quenched activity-based probe and its application in dual color imaging studies of cathepsin S activity localization.

J Am Chem Soc. 2015. 137: 4771-4777.

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Discordance in cathepsin B and cystatin C expressions in bronchoalveolar fluids between murine bleomycin-induced fibrosis and human idiopathic fibrosis

Resp. Res. 2016. In Press

Ketscher A, Ketterer S, Dollwet-Mack S, Reif U, and Reinheckel T.

Neuroectoderm-specific deletion of cathepsin D in mice models human inherited neuronal ceroid lipofuscinosis type 10.

Biochimie. 2016. 122: 219-226.

Tholen M, Wolanski J, Stolze B, Chiabudini M, Gajda M, Bronsert P, Stickeler E, Rospert S, and Reinheckel T.

Stress-resistant Translation of Cathepsin L mRNA in Breast Cancer Progression.

J Biol Chem. 2015. 290:15758-61579.

#### DIPEPTIDYL PEPTIDASE

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DPP9 is a novel component of the N-end rule pathway targeting the Tyrosine Kinase Syk.

eLife. 2016. DOI: 10.7554/eLife.16370

Zhang H, Maqsudi S, Rainczuk A, Duffield N, Lawrence J, Keane FM, Justa-Schuch D, Geiss-Friedlander R, Gorrell MD, and Stephens AN.

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Dipeptidyl peptidase 9 enzymatic activity influences the expression of neonatal metabolic genes.

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#### LEGUMAIN

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## **IMPORTANT PROTEASE PAPERS V**

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Legumain is activated in macrophages during pancreatitis.

Am J Physiol Gastrointest Liver Physiol. 2016. 311: G548-G560.

#### MATRIX METALLOPROTEASES

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A novel mechanism of latency in matrix metalloproteinases.

J. Biol. Chem. 2015. 290: 4728-4740.

Schlage P, Kockmann T, Sabino, F, Kizhakkedathu JN, and auf dem Keller U.

Matrix metalloproteinase 10 degradomics in keratinocytes and epidermal tissue identifies bioactive substrates with pleiotropic functions.

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Active Site Specificity of the Matrix Metalloproteinase Family: Proteomic Identification of 4,300 Cleavage Sites by Nine MMPs Explored with Structural and Synthetic Peptide Cleavage Analyses.

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Degradomic and Yeast 2-Hybrid Inactive Catalytic Domain Substrate Trapping Identifies New Membrane-type 1 Matrix metalloproteinase (MMP14) Substrates: CCN3 (NOV) and CCN5 (WISP2).

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Generation of aggregation prone N-terminally truncated amyloid  $\beta$  peptides by meprin  $\beta$  depends on the sequence specificity at the cleavage site.

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#### **RESEARCH REVIEWS**

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Am J Physiol Gastrointestinal and Liver Physiol. 2016. 310: G234-G239.

#### Seidah NG.

New developments in PCSK9's biology and clinical implications.

Curr. Opin. Lipidol. 2016. 27: 274-281.

Seidah NG, Abifadel M, Prost S, Boileau C, and Prat A.

The proprotein convertases in hypercholesterolemia and cardiovascular diseases: Emphasis on PCSK9.

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Proteomic approaches to uncover MMP function.



## **IMPORTANT PROTEASE PAPERS VI**

#### Matrix Biology. 2015. 44-46:232-238

#### Klein T, Viner R, and Overall CM.

Quantitative proteomics and terminomics to elucidate the role of ubiquitination and proteolysis in adaptive immunity.

Philos Trans A Math Phys Eng Sci. 2016. 374: 1-15.

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Detection of protease activity in cells and animals.

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Lecaille F, Lalmanach G, and Andrault PM.

Antimicrobial proteins and peptides in human lung diseases: a friend and foe partnership with host proteases

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Ono Y, Saido TC, and Sorimachi H.

Calpain research for drug discovery: Challenges and potential.

Nat Rev Drug Discov. 2016 In press.

## Special Issue: The rhomboid protein superfamily in development and disease

#### Guest Editor: Sin Urban

Semin Cell Dev Biol. 2016. S1084-9521(16)

#### Urban S.

A guide to the rhomboid protein superfamily in development and disease.

#### Shilo BZ.

Developmental roles of Rhomboid proteases.

#### рр 30209-9.

#### Lastun VL, Grieve AG, and Freeman M.

Substrates and physiological functions of secretase rhomboid proteases.

#### рр 30237-3.

Spinazzi M, and De Strooper B.

PARL: the mitochondrial rhomboid protease.

pp 30238-5.

Lemberg MK, and Adrain C.

Inactive rhomboid proteins: new mechanisms with implications in health and disease.

#### pp 30184-7.

Dogga SK, and Soldati-Favre D.

Biology of rhomboid proteases in infectious diseases.

pp 30258-0.

#### Bondar AN.

Biophysical mechanism of rhomboid proteolysis: setting a foundation for therapeutics.

pp 30290-7.

#### Strisovsky K.

Rhomboid protease inhibitors: Emerging tools and future therapeutics.

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## Special Issue: Proteolytic networks across cellular boundaries (IPS 2015 Special Issue)

#### Guest Editorial: Judith Clements

Biological Chem. 2016. 397(9)

#### Nagase, H.

A personal journey with matrix metalloproteinases

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#### Murray, AS, Varela, FA, List, K

Type II transmembrane serine proteases as potential targets for cancer therapy

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Kanatsu, K, Tomita, T.

Membrane trafficking and proteolytic activity of  $\gamma\mbox{-secretase}$  in Alzheimer's disease

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#### Wilson, CH. Zhang, HE Gorrell, MD. Abbott, CA.

Dipeptidyl peptidase 9 substrates and their discovery: current progress and the application of mass spectrometry-based approaches

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Schmidt, F. Müller, M. Prox, J. Arnold, P. Schönherr, C. Tredup, C. Minder, P. Ebsen, H. Janssen, O. Annaert, W. Pietrzik, C. Schmidt-Arras, D. Sterchi, EE. Becker-Pauly, C.

Tetraspanin 8 is an interactor of the metalloprotease meprin  $\boldsymbol{\beta}$  within tetraspanin-enriched microdomains



## **IMPORTANT PROTEASE PAPERS VII**

#### pg 857

O'Donoghue, AJ. Ivry, SL. Chaudhury, C. Hostetter, DR. Hanahan, D. Craik, CS.

Procathepsin E is highly abundant but minimally active in pancreatic ductal adenocarcinoma tumors

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Merkulova, Y. Shen, Y. Parkinson, Leigh G. Raithatha, SA. Zhao, H. Westendorf, K. Sharma, M. Bleackley, RC. Granville, DJ.

Granzyme B inhibits keratinocyte migration by disrupting epidermal growth factor receptor (EGFR)-mediated signaling

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Burgener, SS. Baumann, M. Basilico, P. Remold-O'Donnell, E. Touw, IP. Benarafa, C.

Myeloid conditional deletion and transgenic models reveal a threshold for the neutrophil survival factor Serpinb1

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Probing catalytic rate enhancement during intramembrane proteolysis

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Human 20S proteasome activity towards fluorogenic peptides of various chain lengths

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- Spotlight Sessions on protein degradation and proteolysis research
- Oral presentations available (submit abstracts by November 17, 2016)
- 300 travel awards for student/postdoc first author presenters
- Questions? Contact Evette Radisky, Mayo Clinic (radisky.evette@mayo.edu)

#### POSTDOCTORAL POSITION

#### IN PROTEASE BIOCHEMISTRY AND INHIBITION HARVARD MEDICAL SCHOOL

NIH-funded project on the biochemistry and pharmacological modulation of y-secretase, a membraneembedded protease complex central to Alzheimer's disease, cancer biology, and developmental biology. Applicants must have a strong record of achievement in protein chemistry, enzymology and pharmacology. Experience in molecular and cell biology is also required. Candidates must be highly experienced in these specific areas, as demonstrated by journal publications. Send CV and contact information for 2 references to: Michael Wolfe or Dennis Selkoe, Center for Neurologic Diseases, Brigham and Women's Hospital and Harvard Medical School, 4 Blackfan Street, Boston, MA **02115** or e-mail to: mswolfe@partners.org or dselkoe@partners.org.

#### What is on the cover?

A near atomic structure of the active human apoptosome (5JUY.PDB) Tat Cheung Cheng, Chuan Hong, Ildiko V Akey, Shujun Yuan, Christopher W Akey. eLife 2016; 5:e17755

In response to cell death signals, an active apoptosome is assembled from Apaf-1 and procaspase-9. The structure of the active human apoptosome was determined by cryo-EM.





### Post-Doctoral Position available in EXTRACELLULAR MATRIX and MOONLIGHTING PROTEASE DEGRADOMICS: IDENTIFICATION AND CHARACTERISATION OF NOVEL PROTEASE SUBSTRATES

#### OVERALL LAB, University of British Columbia, Vancouver BC, Canada.

A postdoctoral position is available for a candidate to study novel substrates of proteases, in particular matrix metalloproteinases (MMPs). The OVERALL LAB has pioneered a number of cutting edge proteomics methods (degradomics: including **TAILS** (Nature Biotechnology, Nature Protocols), **C-TAILS** (Nature Methods, Nature Protocols), **PICS** (Nature Biotechnology), **TopFIND** database (Nature Methods)) for the analysis of protein termini and identification of protease substrates in healthy and diseased tissue (e.g. arthritis (Cell Reports), skin inflammation (Science Signaling), innate and acquired immunity (Nature Communications), and viral infection (Nature Medicine)). Moonlighting proteins that have canonical roles inside the cell that are secreted by non-conventional means are a priority. The candidate will elucidate the roles of these novel and unexpected substrates in cell regulation before and after proteolytic processing. Thus we will decipher how tissue pathologies, particularly inflammatory diseases, autoimmunity and immunodeficiencies, and cancer, are driven by proteases (*e.g.* MMPs, ADAMs, ADAMTS, cathepsins, viral proteases) and bioactive substrates that modulate signaling feedback loops with emphasis on connective tissues, the extracellular matrix and immune cells.

The candidate should have a practical background in protease biology, extracellular matrix biology, inflammation, and immunobiology. Experience in a variety of biological systems is an asset as the candidate will use biochemical methods, mammalian cell tissue culture and murine models to elucidate the roles of moonlighting proteins and the effects of proteolytic processing as well as determining whether cleavage creates new functions. Biological assays will include angiogenesis, chemotaxis, cell migration and invasion assays, proinflammatory cytokine production. Further characterization of the significance of proteolytic processing *in vivo* will be determined by developing selected reaction monitoring (SRM) assays and neoepitope antibodies to establish the prevalence of proteolytic processing in healthy and inflamed/diseased tissues using mass spectrometry, immunohistochemistry and by developing ELISA-type assays. It is anticipated that some of the identified proteoforms will be useful as biomarkers for disease diagnosis. A significant portion of the lab is dedicated to degradomics, utilizing and developing liquid chromatography tandem mass spectrometry techniques (LC-MS/MS). Thus there is the opportunity for the candidate to use proteomics to address the roles of moonlighting substrates and the applicant will be trained in this.

The lab is situated in the Centre for Blood Research (http://cbr.ubc.ca/) in the Life Sciences Centre (http://lsi.ubc.ca/) at the University of British Columbia in Vancouver.



Apply to chris.overall@ubc.ca, www.clip.ubc.ca

## Save the date for the IPS 2017 General Meeting



## 10th GENERAL MEETING OF THE INTERNATIONAL PROTEOLYSIS SOCIETYBanff Conference Centre, CANADA28 Oct.-1 Nov. 2017

Training workshops and seminars

27-28 Oct. 2017

It is our pleasure to invite you to the 10<sup>th</sup> General Meeting of the International Proteolysis Society from 28 October to 1 November 2017, and early training workshops on the 27-28 October. The meeting will be held at the Kinnear Centre in Banff, in the middle of the Canadian Rockies. It is a splendid venue to be inspired, discuss with colleagues, forge new collaborations and see the best science from the proteolysis field.

Of course we will mark the 10<sup>th</sup> anniversary of our meeting in a special way! We look forward to seeing you in Banff, Alberta. Cowgirl or cowboy boots, belt buckle and bolo tie optional but highly recommended! We'll provide the hat!



Joanne, Jean-Bernard and Chris

### www.IPS2017.org

Meeting topics (preliminary)

- Proteases in Immunity
- Protease Systems Biology
- Biologicals for Therapeutic Intervention
- Proprotein convertases and Metabolism
- Cancer and Metastasis
- Proteasome and ubiquitination systems
- Intracellular Proteolytic Systems
- Proteases in Developmental Biology
- Viral Proteases
- Membrane-Associated Proteolysis
- Proteases in bacteria, fungi and elsewhere

### Save the date for the IPS 2017 General Meeting



## ISK 2017

7 TH INTERNATIONAL SYMPOSIUM ON KALLIKREINS AND KALLIKREIN-RELATED PEPTIDASES

### 26 September – 29 September 2017 TOURS, FRANCE

*Venue: Université F. Rabelais, 60 rue du Plat d'étain, Tours.* 



#### Local Organizing Committee:

Dr. Yves Courty (Chair) Phone: +33 2 47 36 60 50 E-mail: courty@univ-tours.fr Dr. Agnès Petit-Courty E-mail: agnes.petit@univ-tours.fr

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A. Scorilas (Greece); M. Schmitt (Germany); G. Sotiropoulou (Greece); G. Yousef (Canada).





## SAVE THE DATE

### Symposium on

## Proteases and the Tumori Microenvironment

This will be the highlight of the year for the Proteases and Tumori Microenvironment aficionados! Save the date now!

### 24 - 26 July 2017 Monash University Prato Centre, Prato, Italy

Organisers: Gail Risbridger James Whisstock Judith Clements

## www.ProteasesinTiME.org

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Margarete Heck

University of Edinburgh Margarete.heck@ed.ac.uk



Novo Nordisk hrse@novonordisk.com

raisk.com

Judith Clements Queensland University of Technology

j.clements@qut.edu.au

www.protease.org



Taisuke Tomita University of Tokyo taisuke@mol.f.u-tokyo.ac.jp





Maria Luiza Vilela Oliva Universidade Federal de São Paulo olivamlv@gmail.com

#### Ex Officio



Bob Lazarus Genentech Iazarus.bob@gene.com

#### Webmaster



Christian Sommerhoff Ludwig Maximilians University sommerhoff@med.uni-muenchen.de Sheena McGowan d Monash University sheena.mcgowan@monash.edu



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### The International Proteolysis society is always keen to keep in touch! Email us today or find us on





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